FAGOPYRUM

NOVOSTI O AJDI – NAJNOWSZE INFORMACJE O GRYKE BUCKWHEAT NEWSLETTER – БЮЛЛЕТЕНЬ »ГРЕЧИХА« NOVOSTI O HELJDI – LES ANNALES DE SARRASIN **愛え: समादार – पत्रक ソバ ニューズレター**

KAZALO – CONTENTS

- 3 R. AMAROWICZ, Ł. FORNAL: Characteristics of buckwheat grain mineral components and dietary fiber
- 7 I. TAHIR, S. FAROOQ: Occurrence of phenolic compounds in four cultivated buckwheat with particular reference to grain quality
- 9 S. FAROOQ, I. TAHIR: Comparative study of some growth attributes in buckwheats cultivated in Kashmir
- 13 B. BOHANEC: Improvements in buckwheat micropropagation procedures
- 16 L. KAJFEŽ-BOGATAJ: Light and temperature dependence of net photosynthesis for buckwheat
- 19 P. NARAIN: Studies on buckwheat improvement
- 20 M. R. BHAT: Aphelinus kashmiricus Hyat a parasite of Aphis gossypii Glover (Melon Aphid) in Kashmir
- 21 L. KAJFEŽ-BOGATAJ: Study on the development and contributions of different plant organs to buckwheat dry matter production
- 24 M. RUSZKOWSKI: IBRA information
- 26 Contents of Proc. of 3rd Int. Buckwheat Symposium, Pulawy, Poland 1986
- 28 SEZNAM NASLOVOV MAILING LIST

FAGOPYRUM (NOVOSTI O AJDI), zvezek 7, Ljubljana 1987:

Izdajatelji:

VTOZD za agronomijo, VDO Biotehniška fakulteta, Krekov trg 1, 61001 Ljubljana,

Zveza društev genetikov Jugoslavije in Društvo genetikov Slovenije, Ljubljana.

Za publikacijo odgovarja: Ivan Kreft.

Opremil: Sandi Radovan.

Ofset tisk: Birografika BORI, Ljubljana, Titova 64 (600 izvodov). Cena izvoda 400 din.

V publikacijo so vključeni povzetki objavljenih člankov in predhodne informacije o še neobjavljenih rezultatih raziskavo ajdi. Namenjena je za uporabo na Biotehniški fakulteti in za brezplačno medsebojno izmenjavo za publikacije drugih inštitucij. Raziskovalci in knjižnice prejmejo po en izvod brezplačno.

Po mnenju republiškega komiteja za kulturo z dne 14. 5. 1986 je publikacija oproščenatemeljnega davka od prometa proizvodov.

FAGOPYRUM (BUCKWHEAT NEWSLETTER), Vol. 7, Ljubljana 1987.

Published under the auspices of the International Buckwheat Research Association (IBRA), of Union of Yugoslav Genetic Societies and of the Genetic Society of Slovenia.

Editor: Ivan KREFT (Biotehniška fakulteta, Univerza Edvarda Kardelja, 61001 Ljubljana, Yugoslavia).

Advisory committee: Taiji ADACHI (Fac. of Agriculture, Miyazaki University, Kumano 7710, Miyazaki 889-21, Japan), N. V. FESENKO (All-Union Research Institute of Legume and Groat Crops, Orel, USSR), Toshiko MATANO (Fac. of Agriculture, Shinshu University, Ina, Nagano-Ken, Japan), Takashi NAGATOMO (7075 Funahiki, Kiyotake-cho, Miyazaki, 889–16, Japan), V. P. SINGH (School of Studies in Botany, Vikram University, Ujjain 456 010, India).

Published by: VTOZD za agronomijo, genetika, Biotehniška fakulteta, Univerza Edvarda Kardelja v Ljubljani, Krekov trg 1, 61000 Ljubljana, Jugoslavija.

Materials, prepared for direct photocopying, received not later than March 10, next year, will be included in next Volume. No special priority is given to any language. The publication willbe distributed free (the exchange of publications is appreciated) to scientists, libraries etc., who would like to be included on the mailing list.

FAGOPYRUM (Buckwheat Newsletter) is open to everyone interested in buckwheat and will cover all aspects of buckwheat research: genetics, cytology, breeding, cultivation, nutrition, utilization, biochemistry and other. Scientific papers, reviews, research notes on work in progress or on results not yet published: comments and speculations related to buckwheat; list of stock materials wanted or available; lists of names, addresses and field of work of scientists who have expressed the desire to receive the Newsletter; lists of publications which are related to buckwheat and which have appeared during preceding years; announcements concerning the promotion of research on buckwheat (workshop, symposia, and so on): abastracts and/or contents of published papers/books on buckwheat; bibliographies and other information related to buckwheat or buckwheat research will be published. In order to facilitate the elaboration of the bibliography scientists are asked to send reprints of their own publications to the editor of Fagopyrum.

Characteristics of buckwheat grain mineral components and dietary fiber

Ryszard Amarowicz and Łucja Fornal

Chair of Human Nutrition and Institute of Engineering and Biotechnology, University of Agriculture and Technology, 10-718 Olsztyn, Poland

In search for co called healthy food sources of high nutritive value chemists and technologists turned to buckwheat. The result of investigations is chemical and physical-chemical characteristics buckwheat protein (14, 15, 18) and determined high nutritive value (11); its available carbohydrates (19) and lipids (20) are better and better described. Relatively little research has been devoted to mineral components and nonavailable carbohydrates. Acc. to Ruszkowski and Ruszkowska (16), apart from phosphorus and iron, buckwheat grain contains potassium, zink, and some small amounts of boron, iodine, nickel and cobalt. Acc. to Trzebska-Jeske (21) buckwheat groats contain potassium, boron, iron, copper, chromium, zink, cobalt, nickel, and also calcium, magnesium, phosphorus and sodium. Similar chemical composition of mineral salts was stated by Zalesskaja and Mielnikow (23) though it varied in quality from other data (21). Buckwheat grain INQ for mineral components was acc. to Amarowicz and Fornal (1) P = 3.10, Fe - 1.91, Mg - 4.62, Zn - 1.98. These figures, greater than 1.00, prove that buckwheat groats can satisfy demands of mineral components in a higher degree than his energetical demands. There is no unanimous view on the qualitative and quantitative mineral composition accompanying structural carbohydrates. Their definition of the last decade takes into account fractions of hemicelluloses, celluloses and lignins and of other substances, which do not undergo enzymatic hydrolysis. The content of structural carbohydrates in buckwheat was acc. to the 1960s definition, or so colled crude fiber, 12-18 % d.m. Crude fiber content in the seed coat was determined as ab. 50 %, and in groats – as ab. 1 % (12).

Above nonunanimous statements made us undertake research aiming at determining the content of selected mineral components in buckwheat grain, flour, groats and seed coat, as well as at determing dietary fiber content and its fractions in the same material.

Materials and methods

Seed coat, groats and flour obtained from trade buckwheat grain were investigated. Material was reduced in size to $> 315 \ \mu m$ granulated product.

Mg, Fe, Zn, Cu and Mn content was determined with AAS, K with flame photometry (17). Nitric acid and perchloric acid mixture (4 : 1, v v) was applied for mineralization. The IL 353 spectrophotometer and Flapho IV flame photometer were used. After 1.5 h preheating at water boiling temperature dietary fiber was determined acc. to Hellendoorns method (17), and acid detergent fiber – acc. to Van Soests method (22).

Dietary fiber = hemicelluloses + celluloses + lignins Acid detergent fiber = celluloses + lignins

Discussion of results

Mineral components content in the examined material was characterized by a diverse distribution (Tab. 1). Compared to cereal grain buckwheat seed coat is rich only in iron and manganese, while compared to whole buckwheat grain it contains less zinc, copper and potassium, and less manganese. Hence it can be assumed that buckwheat endosperm is very rich in mineral salts and removing seed coat does not affect qualitative-quantitative composition of flours mineral salts.

Compared to flour buckwheat groats were found to contain less zinc, manganese and iron, and more potassium and magnesium. This points to the possibility of migrating of these elements from seed coat to the possibility of migrating of these elements from seed coat to endosperm during hydrothermic treatment. Magnesium and potassium content in the investigated material is higher than in cereal grain and its products.

From the nutritional point of view content of Mg, an element esential for the right course of many biochemical processes in human organism, is especially valuable.

Results obtained for buckwheat grain are for Mg, K and Cu lower, and for Zn and Mn higher than those quoted by Javornik (12). Considerably higher amounts of Mg (362–365 mg %) was found in buckwheat by Kusano et al (13). Bubicz and Sklarz (4) and Bubicz et al. (3) observed much lower level of Mg (62–82 mg % for the Emka veriety and 93–109 mg % for the Hruszowska variety). They also noticed similar contents of Zn, Cu, Mn and Fe, and higher K level than in the present work. Investigating buck-

wheat groats Trzebska – Jeske et al. (21) presented following values (mg 100 g d.m.): Mg – 216, Fe – 2.82, K – 475, Cu – 0.40, Zn – 3.50, Mn – 2.04. In Dietrych-Szóstaks and Ploszynskis work (6) Mg content in groats was 200-300 mg/100 g d.m. and on the way of dehulling.

The problem of dietary fiber has been the object of interest of many chemists and nutritionists for the last decade. Developing new methods of determining contributed to investigating quality and quantity composition of dietary fiber in cereal grain and in other seeds. The results of our investigations characterize fractional composition of dietary fiber of buckwheat grain and its products (Tab. 2).

Dietary fiber content in groats and flour was much lower than in buckwheat grain while seed coat was found to contain 3 times more fiber than the grain. In fractional composition of dietary fiber in the examined material there is a noticeably higher amount of lignins and celluloses in comparison to hemicelluloses (Fig. 1). The seed coat fiber contained the greatest amount of celluloses and lignins, and this relation brings upon seed coat nonconsumption utilization (10). An interesting phenomenom is the presence of hemicelluloses in buckwheat groats. It can be assumed to be result of celluloses changing during hydrothermic process. It is quite possible that forms of higher solubility remained in acid solution of detergent and eventually the results of hemicelluloses content were overestimated.

Dietary fiber content determined in our investigations is higher then it is in so called crude fiber (2). For various buckwheat varieties crude fiber contents are following: 15.17-15.79 % (3), 13.71-15.21 % (4), 17.79 and 12.72 % d.m. (9). In buckwheat groats they were 1.18 % d.m. (10), 0.29 % d.m. (9), and 0.24-0.60 % d.m. (7), and in seed coat -44.49 % d.m. (8). Bulinski et al. (5) found in buckwheat groats higher contents of dietary fiber, i.e. 8,38-10.21 % d.m. Also Dietrych-Szóstak and Ploszynski (6) showed higher part of celluloses and hemicelluloses than the present results. The divergences may be result not only from applying different analytical methods but also from the range of buckwheat varieties.

Summing up the present results it is worth stressing the typical quality and quantity composition of mineral components in buckwheat grain and its products whereas the composition of dietary fiber being dominated by celluloses and lignins. In cereal grain the main fraction of the fiber are hemicelluloses making 60 % of cereal bran.

Material	К	Mg	Mn	Fe	Zn	Cu
Buckwheat grain	244,1	168,6	5,44	4,82	3,40	0,59
Flour	299,4	160,8	2,22	5,26	3,72	0,53
Seed coat	248,0	120,6	13,09	5,61	2,49	0,49
Groat	301,5	172,9	1,53	3,69	2,96	0,79

Table 1. Mineral components content in buckwheat and its products (mg/100 g)

Table 2. Dietary fiber content and its composition in buckwheat grain and its products (% d.m.)

			1014-00 M		
Material	Dietary fiber	detergent fiber	Hemi- celluloses	Celluloses	Lignins
Buckwheat grain	24,75	20,39	4,36	9,81	10,58
Flour	3,94	3,98	0,00	2,34	1,64
Seed coat	80,31	61,98	18,33	29,93	32,05
Groat	4,51	2,29	2,22	1,81	0,48

4

Fig. 1. Composition of dietary fiber in buckwheat grain and its products

grain



Buckwheat



Flour









Hemicelluloses [[]]Celluloses 😑 Lignins

References

- 1. Amarowicz R., Fornal Ł. (in the press): Wskaźnik wartości zywieniowej produktów zbozowych. Przegl. Gastronom.
- Asp N.G. (1978): Critical evaluation of some suggested methods for assay of dietary fiber. J. Plant Technol. (3(1), 21-25.
- Bubicz M., Korzeń A., Korzeń S. (1986): Effect of increasing nitrogen rates on the chemical composition of buckwheat seeds. Proceedings of the 3rd International Symposium on Buckwheat, Pulawy, Poland, 7-12 July 1986.
- Bubicz M., Sklarz J. (1986): Effect of selected mirconutrients on the growth, yield, and chemical composition of buckwheat. Proceedings of the 3rd International Symposium on Buckwheat, Pulawy, Poland, 7-12 July 1986.
- Buliński R., Bloniarz J., Kot A., Szydlowska E., Wyszogrodzka L. (1986): Badanie zawartości blonnika pokarmowego w kaszach, ryzu, platkach zbozowych, makaronach, pieczywie trwalym oraz w niektórych odzywkach dla dzieci. Roczn. PZH 37(1), 48-51.
- Dietrych-Szóstak D., Ploszyński M. (1986): Chemical composition and feeding value of buckwheat hulls and harvest residues. Proceedings of the 3rd International Symposium on Buckwheat, Pulawy, Poland, 7-12 July 1986.
- Dietrych-Szóstak D., Ploszyński M. (1986): Some aspects of the nutritional value of light and dark buckwheat groats. Proceedings of the 3rd International Symposium on Buckwheat, Pulawy, Poland, 7-12 July 1986.
- 8. Dietrych-Szóstak D., Ploszýnski M. (1986): Some analytical data on the biological quality of buckwheat groats and hulls obtained from the seeds of cv. Hruszowska. Fagopyrum 6, 21.
- 9. Eggum B. O., Kreft I., Javornik B. (1981): Chemical composition and protein quality of buckwheat (Fagopyrum esculentum Moench). Qual. Plant. Plant Foods Hum. Nutr. 30, 175-179.
- 10. Javornik B., Eggum B.O., Kreft I. (1981): Studies on protein fractions and protein quality of buckwheat. Genetika 13 (2), 115-121.
- 11. Javornik B. (1983): Nutritional quality and composition of buckwheat proteins. Proceedings of the 2nd International Symposium on Buckwheat, Miyazaki, 1983.
- Javornik B. (1986): Buckwheat in human diets. Proceedings of the 3rd International Symposium on Buckwheat, Pullawy, Poland, 7-12 July 1986.
- 13. Kusano T., Chiue H., Ikeda K., Arihara M. (1983): Nutritive components in autotetraploid buckwheat. Proceedings of the 2nd International Symposium on Buckwheat, Miyazaki, 1983.
- Pomeranz Y., Robbins G.S. (1972): Amino acid composition of the buckwheat proteins. J. Agric. Food Chem. 20(2), 270-274.
- 15. Pomeranz Y. (1973): A review of proteins in barley and buckwheat. Cereal Sci. Today 18(9), 310-315.
- 16. Ruszkowski J., Ruszkowska B. (1981): Gryka, PWRiL, Warszawa.
- 17. Rutkowska U. (1981): Wybrane metody badania skladu i wartości odzywczej zywności. PZWL, Warszawa.
- 18. Soral-Śmietana M. (1984): Bialka gryki. Postęppy Nauk Roln. 3, 35-46.
- Soral-Śmietana M., Fornal Ł., Fornal J. (1984): Characteristics of of buckwheat grain starch and the effect of hydrothermal processing upon its chemical composition, properties and structure. Starch-Starke 36(5), 153-158.
- 20. Soral-Śmietana M., Fornal Ł., (1984): Characteristics of lipids in buckwheat grain and isolated starch and their changes after hydrothermal processing. Die Nahrung 28(5), 483-492.
- 21. Trzebska-Jeske I., Nadolna I., Rutkowska U., Secomska B. (1973): Wplyw obróbki mechanicznej na wartość Odzywczą kasz produkowanych w kraju. Roczn. PZH 24(6), 717–724.
- Walicka E. (1978): Zastosowanie krajowych detergentów w metodzie Van Soesta oznaczania wlóknika i ligniny. Roczn. Nauk Roln. B, 94-I, 140-147.
- 23. Zalesskaja E. W., Mielnikow E. M. (1978): Izmienije mineralnogo sostawa jadricy pri gidrotermiczeskoj obrabotke. Izviestia Vyssich Uczebnych Zaviedienij. Piszczewaja Technologia, 7, 43–44.

Izvleček

V zrnju, moki in ajdovi kaši so bile analizirane mineralne snovi in vlakna. S prehranskega vidika je pomembno, da je v zrnju ajde več magnezija kot v zrnju drugih žit. V ajdi je tudi razmeroma veliko vlaken, zlasti celuloze in lignina, medtem ko med vlakni žit prevladujejo hemiceluloze.

Occurrence of phenolic compounds in four cultivated buckwheats with particular reference to grain quality.

Inayatullah Tahir and Sikandar Farooq

Department of Botany, University of Kashmir, Srinagar-190006; Kashmir, India.

Key words: buckwheat, phenolic compounds, species.

Buckwheat (Fagopyrum sp., Fam. Polygonaceae) are economically important primarily due to their nutritious grains (Tahir and Farooq [10]), besides foliage being used as a green vegetable (Narain [6]). Four speices of Fagopyrum viz. F. esculentum Moench, F. sagittatum Gilib., F. tataricum Gaertn. and a new species F. kashmirianum Munshi [5] grow in mixed stands at various high altitude areas of Kashmir as reported by Tahir and Farooq [9]. The present communication reports the occurrence of some phenolic compounds in foliage and in grains of the four species with a view for subsequent improvement of grain quality.

The phenolic compounds were separated on paper chromatograms from ethanolic extracts according to Margna and Margna [4], using upper phase of isoamyl alcohol-petrol ether, acetic acid and water (3:1:1:1) as the solvent system. The spots were located by viewing the chromatograms directly under UV light using a Toshniwal UV lamp with emission maximum at 360 nm. Besides the chromatograms were also viewed under UV light after exposing them to NH₃ vapour or spraying them with AlCl₃.

In the leaf tissue fractions two compounds separated each at $R_f 0.33$ and 0.94 were located on chromatograms in each of the four buckwheat species; the former spot corresponded to rutin due to its typical UV light absorption characteristics, the latter could not be characterised. The present finding is commensurate with the widely known occurrence of rutin in the leaves of *F. esculentum* and *F. tataricum* (Singh and Atal [8]).



Fig. 1. UV-visible light absorption spectrum of the flavonoid compound in the groat fraction of grains of *Fagopyrum kashmirianum* located at $R_f 0.58$ on paper chromatograms.

Fig. 2: Bathochromic shift observed in A_{max} of the flavonoid compound detected in the groat fraction of the grains of *Fagopyrum kashmirianum* at $R_f 0.58$ on paper chromatograms.

The chromatograms of hull fraction of grains, when viewed under UV light showed a prominent spot of rutin at $R_f 0.36$ in each of the four species, however, the spot was inconspicuous in case of *F. kashmirianum*. The chromatograms of the groat fraction, when viewed under UV light and exposed to NH_3 vapours showed the presence of two spots possessing yellowish absorbance; separated each at $R_f 0.58$ and 0.83 in each *F. sagittatum*, *F. tataricum* and *F. kashmirianum*, however, both the spots were absent in *F. esculentum*. When the chromatograms of the groat fraction were sprayed with AlCl₃ and observed under UV light, the spot at $R_f 0.58$ gave intense yellow fluorescence and that at $R_f 0.83$ gave faint yellow fluorescence.

The compound separated at $R_f 0.58$ in the groat fraction has tentatively been characterised. The clear eluate of the flavonoid spot in 95 % ethanol was subjected to spectrophotometry in the UV visible range (170-500 nm) in a Pye Unicam model Sp8-100-UV spectrophotometer. Study of the absorption spectra of the compound showed that the compound has its A_{max} at 372.5 nm (Fig. 1). Addition of a drop of 5 % alcoholic AlCl₃ produced a 56 nm bathochromic shift in A_{max} (Fig. 2). These characteristics closely correspond to those of the flavonol quercetin which has its A_{max} at 374 nm and produces a 70 nm bathchromic shift with AlCl₃ (Harborne [2]). The present findings are commensurate with those of Sato et al. [7], who have isolated and identified quercetin from the grains of tartary buckwheat.

The better palatability of the grains of F. esculentum has been attributed to a low content of phenolics both in hull and groat fractions as compared to the other three species [10]. Besides, the low protein digestibility of buckwheat grains has been attributed to a high content of phenolics by Eggum et al. [1] and Javornik et al. [3]. Inspite of this disadvantage the buckwheat species high in phenolic content predominate in this region where these are cultivated in mixed stands [9]. Surprisingly the phenol rich grains belong to the species which are self compatible (unpublished results of the authors). The grain quality of the three self compatible buckwheats could be improved by testing various genetical procedures.

References

- 1. Eggum B.O., Kreft I., Javornik B. (1981): Chemical composition and protein quality of buckwheat (*Fagopyrum esculentum*). Qual. Plant. Plant Foods Hum. Nutr. 30: 175–179.
- 2. Harborne J. B. (1973): Phytochemical Methods. Chapman and Hall Ltd London, pp. 33-80, 166-204.
- 3. Javornik B. Eggum B.O. Kreft I. (1981): Studies on protein fractions and protein quality of buckwheat, Genetika 13: 115-121.
- Margna U., Margna E. (1969): A suitable chromatographic method for the quantitative assay of rutin and flavone- C-glycosides in buckwheat seedlings. Eesti NSV Tead Akad. Toim Biol. 18: 139-150.
- 5. Munshi A.H. (1982): A new species of *Fagopyrum* from Kashmir Himalaya. J. Econ. Tax. Bot. 3: 627-630.
- 6. Narain P. (1983): Buckwheat in India. Fagopyrum 3:7-11.
- 7. Sato H., Furakawa E., Sakamura S. (1980): Isolation and identification of flavonoids in tartary buckwheat seed (*Fagopyrum tataricum* Gaertn.). Nippon Nogeikagaku Kaishi 54: 275–277.
- Singh A., Atal C.K. (1982): Cultivation of buckwheat in the plains as a raw material for rutin. Cultivation and Utilization of Medicinal Plants, Publication and Information Directorate, CSIR, New Delhi, pp 337-341.
- 9. Tahir I., Farooq S. (1983): Growth and yield characteristics of some buckwheats (Fagopyrum Gaertn.) from Kashmir, Geobios 10: 193–197.
- 10. Tahir L., Farooq S. (1985): Grain composition in some buckwheat cultivars (*Fagopyrum* Spp.) with particular reference to protein fractions. Qual. Plant. Plant Foods Hum. Nutr. 35: 153-158.

Izvleček

Pri štirih vrstah rodu *Fagopyrum* so bile analizirane fenolne snovi. V listju štirih vrst sta bili ugotovljeni s pomočjo kromatografije dve fenolni spojini, od katerih je ena rutin, druga pa je še neidentificirana. Raziskava listov je pomembna, saj se ajdovo listje uporablja kot zelenjavo. V luskah zrn je izrazito prisoten rutin. Kromatogrami kaše kažejo razen pri F. esculentum dve rumenkasti mesti.

Comparative study of some growth attributes in buckwheats (Fagopyrum sp.) cultivated in Kashmir

Sikandar Farooq and Inayatullah Tahir

Department of Botany, University of Kashmir, Srinagar-190006, India.

A comparison has been made of some growth attributes of four species of *Fagopyrum* viz. *F. esculentum* Moench, *F. sagittatum* Gilib., *F. tataricum* Gaertn, and *F. kashmirianum* Munshi. *F. esculentum* attains maximum dry weight, leaf area and height earlier as compared to the other three species, among which *F. tataricum* attains maximum values for these parameters at a later stage. *F. tataricum* seems to possess a higher productivity potential owing to its longer growth span. *F. esculentum* shows higher values for net assimilation rate (NAR) and relative growth rate (RGR). The biomass duration (BMD) and leaf area duration (LAD) was found to be maximum in *F. tataricum*, however, *F. esculentum* attained peak values for BMD and LAD at an earlier stage as compared to the other three species.

Introduction

Buckwheats (Fagopyrum sp., Fam. Polygonaceae) are economically important due mainly to their protein rich grains, hardiness of plants, short growth span, besides foliage being used as a green vegetable (Tahir and Farooq, 1983, 1985; Narain, 1983). Four species of Fagopyrum viz. F. esculentum Moench, F. sagittatum Gilib., F. tataricum Gaertn. and F. kashmirianum Munshi have been reported to grow in mixed stands at certain high altitude areas of Kashmir (Tahir and Farooq, 1983).

Only a little information is asvailable on various phytometric characteristics of buckwheats (Midorikawa, 1957; Ivaki, 1959; Tahir and Farooq, 1983; Kajfež-Bogataj and Knavs, 1985; Kajfež-Bogataj and Gaberščik, 1986). The role of phytometric investigations in studies of plant stand architecture is very important for studying productivity and especially for modelling of canopy net photosynthesis (Ross, 1981). Growth analysis helps to elucidate the interaction between the crop and its physical environment. The study shall enable the modelling of buckwheat production for various genotypes with different growth patterns.

P	Species					
Parameter	F. escul.	F. sagit.	F. kash.	F. tata.		
Total dry wt. g plant ⁻¹	5.41 (10) ±1.37	6.16 (15) ±2.27	7.21 (14) ±1.49	17.51 (12) ±2.89		
Root dry wt. g plant ⁻¹	1 (10)	0.810(11)	0.780(11)	1.790 (12)		
Leaf dry wt. g plant ⁻¹	1.60 (9)	1.70 (10)	3.10(10)	3.39 (12)		
Dry wt. of branches						
g plant ⁻¹	5.14	4.70 (10)	5.90 (10)	9.94 (12)		
Percent moisture						
content plant ⁻¹	84.48 (5)	85.78 (6)	84.69 (5)	87.73 (5)		
Leaf area $cm^2 plant^{-1}$	$250(7) \pm 45.28$	321.51 (11) ±47.15	5 483.57 (10) ± 87.4	$0695.81(12) \pm 87.90$		
Height cm plant ⁻¹	$104.00(9) \pm 6.44$	109.15 (14) ±8.19	110.40 (11) ±8.03	143.40 (14) ±3.21		

Table 1. Maximum values of growth parameters recorded for four species of Fagopyrum.

Figure in parentheses represent growth period in weeks.

Material and methods

Seeds of the four species of *Fagopyrum* viz. *F. esculentum*, *F. sagittatum*, *F. tataricum* and *F. kashmirianum* were separately sown in beds $(3 \times 1 \text{ m})$ randomised into three blocks for two successive generations i.e. 1980 and 1981. The distance between the rows was 10–15 cm and within the rows the seeds were sownt at a distance of 5 cm. The seeds were sown at a depth of 5–7 cm. Soil moisture levels were generally maintained above 50 % of field capacity. To support optimal growth, soil dressing of fertiliser was applied twice, i.e. a week prior to sowing and at 4 weeks growth. The doses of the fertilizer applied were as follows: N 57 kg/ha, P 38 kg/ha and K 40 kg/ha. The experiment was terminated at 15 weeks growth. At each weekly interval 5 plants were sampled at random. Leaf area was estimated gravimetrically by the method described by Šesták *et al.* (1971). For the estimation of dry weight the fresh material was dried in an oven at 70°C for 48 hours. Different growth parameters were computed on the basic of formulae described by Evans (1972) and Hunt (1978). For finding net assimilation rate on leaf dry weight basis formula of Blackman and Black (1965) was used. For finding leaf area partitioning coefficient formulation of Potter and Jones (1977) was followed.

Results and discussion

Maximum dry weight, leaf area and height were attained relatively earlier in *F. esculentum* than in the other three species, among which maximum values for these parameters were attained much later in *F. tataricum* (Table 1, Figs. 1 and 2). The dry weight was found to be closely related to leaf area, thus reflecting the influence of the size of the assimilatory apparatus in determining the dry matter production.

The higher dry matter production in *F. tataricum* is commensurate with the highest leaf area in this species. It is obvious that *F. tataricum* possesses a higher productivity potential due to its greater accumulation of total phytomass and its partitioning into different plant parts. A considerably larger proportion of the dry weight was partitioned into the top parts than into roots in plants of each of the four species, making them succeptible to lodging.

Demonster	Species					
Parameter	F. escul.	F. sagit.	F. kash.	F. tata.		
NAR mgcm ⁻² week ⁻¹			Collector of the coll			
(leaf area basis)	13.87(6-7)	9.41 (4-5)	8.42(6-7)	11.37(10-11)		
NAR mg mg ⁻¹ week ⁻¹						
(leaf dry wt. basis)	5.33(6-7)	2.92(6-7)	2.05(3-4)	2.51(5-6)		
LAR $cm^2 mg^{-1}$	0.213(3-4)	0.129(5-6)	0.219(4-5)	0.197(3-4)		
LWR g g^{-1} week ⁻¹	0.646(3-4)	0.650(3-4)	0.636(3-4)	0.596(3-4)		
RGR g g^{-1} week ⁻¹	1.86(3-4)	1.37(3-4)	1.30(3-4)	1.70(4-5)		
RGR Roots	2.24(6-7)	1.61(6-4)	1.31(4-5)	1.90(4-5)		
RGR Leaves	1.28(3-4)	1.36(3-4)	1.23(3-4)	1.48(5-6)		
LAER $cm^2 cm^{-1} week^{-1}$	1.26 (2-3)	1.70 (4-5)	2.05 (3-4)	1.25 (4-5)		
LAPC	1 250 (2 2)	0.101/1.7	0 107 (0 1)	0.000 (0		
cm ² week ⁻¹ /mg week ⁻¹	1.350(2-3)	0.181(4-5)	0.436(3-4)	0.228 (3-4)		
BMD g week ^{-1}	7.53 (9-10)	6.82(10-11)	8.04(10-11)	17.07 (12-13)		
$LAD cm^2 week^{-1}$	219.99 (7-8)	311.31 (10-11)	456.49 (10-11)	659.74 (11-12)		

Table 2. Maximum values of some growth analysis parameters recorded for four species of *Fagopyrum* (For abbreviations see the text).

Figures in parentheses represent growth interval in weeks.

Maximum recorded value for net assimilation rate (NAR) and relative growth rate (RGR) was found to be higher in *F. esculentum* as compared to the other three species; among which *F. tataricum* possessed higher values for NAR and RGR than in *F. sagittatum* and *F. kashmirianum* (Table 2). NAR was found to be closely related to RGR as reported earlier (Singh and Singh, 1982). The RGR of the leaves becomes important as it is the determinant of the time taken for a crop to develop its light intercepting canopy (Evans, 1976). Generally *F. tataricum* and *F. esculentum* possessed a higher RGR of leaves and roots as compared to the other three species. The ratio of photosynthesising to respiring material i.e. leaf area ratio (LAR) was found to be much lower in *F. sagittatum*, however, the ratio of leaf dry weight to whole plant dry weight i.e. leaf weight ratio (LWR) was found to be slightly lower in *F. tataricum* as compared to the other three species.

Leaf expansion per unit dry weight per unit time i.e. leaf area partitioning coefficient (LAPC) was found to be lower in *F. sagittatum*, *F. kashmirianum* and *F. tataricum* which suggests that there is more dry matter available for supporting leaf area expansion in these species. *F. sagittatum* and *F. kashmirianum* possessed a higher leaf area expansion rate (LEAR) compared to that in *F. esculentum* and *F. tataricum*.

Fig. 1: Plants of four species of *Fagopyrum* grown in pot culture. From left to right are arranged pots with plants of *F. sculentum*, *F. sagittatum*, *F. kashmirianum* and *F. tataricum* (photographed at eight weeks growth).



Fig. 2: Plants of four species of *Fagopyrum* grown in pot culture. From left to right are arranged pots with plants of *F. esculentum*, *F. sagittatum*, *F. kashmirianum* and *F. tataricum* (photographed at sixteen weeks growth).

Biomass duration (BMD) i.e. the dry matter production of crop's growing period and leaf area duration (LAD) i.e. the leafiness of crop's growing period was found to be maximum in *F. tataricum* owing to its longer growth span. However, *F. esculentum* attained the maximum values for BMD and LAD at an eralier stage due mainly to its quick growing efficiency as compared to the other three species. LAD thus appears to effectively determine the overall photosynthetic potentiality of the species and inturn determines BMD.

References

Blackman, G.E. and Black, J.N. 1965. Physiological and ecological studies in the analysis of plant environment. An analysis of the effect of seasonal variation in daily light and temperature on the growth of *Helianthus annus* in the vegetative phase. Ann. Bot., *19:* 327–348.

Evans, G.C. 1972. The Quantitative Analysis of Plant Growth. Blackwell Sci. Publ. Oxford.

Evans, L.T. 1976. Physiological adaptation to performance as crop plants. Phil. Trans. R. Soc. Lond. B., 275: 71-83.

Runt, R. 1978. Plant Growth Analysis. Edward Arnold Publishers, Ltd. London.

Ivaki, H. 1959. Ecological study on interspecific competition in a plant community. I. An analysis of growth of competition in mixed stands of buckwheat and green grains. Jap. J. Bot., 17: 120-138.

Kajfež-Bogataj, L. and Gaberščik, A. 1986. Analysis of net photosynthesis response curves for buckwheat, Fagopyrum, 6: 6-7.

Kajfež-Bogataj, L. and Knavs, M. 1985. Studies on the production of dry matter in the community of buckwheat with particular reference to leaf area. Fagopyrum, 5: 7-12.

Midorikawa, B. 1957. The distribution of CO_2 concentration of the air in some plant communities. Jap. J. Ecol., 7: 72-76.

Narain, P. 1983. Buckwheat in India. Fagopyrum, 3: 7-11.

Potter, J.P. and Jones, J.W. 1977. Leaf area partitioning as an important factor in growth. Plant Physiol., 59: 10.

Ross, J. 1981. The radiation regime and architecture of plant stands. Dr. W. Junk Publishers, The Hague, 391 pp.

Šesták, Z., Čatský, J. and Jarvis, P.G. 1971. Photosynthetic Production, Manual of Methods. Dr. W. Junk Publishers, The Hague, 517–555.

Singh, B.G. and Singh, J.N. 1982. Effect of seasonal changes on growth parameters of green gram (Vigna radiata L.) Wilczek. Ind. J. Plant Physiol., 25: 382-389.

Tahir, I. and Farooq, S. 1983. Growth and Yield characteristics of some buckwheats (Fagopyrum Gaertn.) From Kashmir. Geobios, 10: 193–197.

Tahir, I. and Farooq, S. 1985. Grain composition in some buckwheat cultivars (*Fagopyrum* Spp.) with particular reference to protein fractions. Qual. Plant. Plant Foods Hum. Nutr. *35*: 153–158.

Izvleček

Analizirana je bila rast štirih vrst rodu *Fagopyrum. F. esculentum* doseže največjo težo sušine, največjo listno površino in višino rastlin preje kot ostale tri vrste, medtem ko doseže *F. tataricum* maksimalne vrednosti teh parametrov najkasneje. Kot kaže, ima *F. tataricum* največji potencialno možen pridelek zaradi dolge življenjske dobe. *F. esculentum* ima višjo stopnjo neto asimilacije in relativne rasti. Trajanje biomase in trajanje listne površine sta največji pri tatarski ajdi, čeprav pri obojem doseže maksimalne vrednosti najpreje navadna ajda.

Improvements in buckwheat (Fagopyrum esculentum Moench) micropropagation procedures

Borut Bohanec

VTOZD za agronomijo, Biotehniška fakulteta, Univerza Edvarda Kardelja v Ljubljani, 61000 Ljubljana, Jugoslavija

Abstract

With buckwheat micropropagation started from seed, experiments were focused on radicula removement 6 days after in vitro germination followed by successful axillary bud induction, subculturing of single shoots and addition of liquid media instead of subculturing. Cutting of adult plants and induction of young shoots followed by inoculation of young inflorescences, could be a method for successful induction of micropropagation started from adult plants.

Introduction

In vitro procedures for buckwheat micropropagation have been reported in several papers (see references Yamane, 1974; Nešković et al., 1986; Bohanec, 1986). In the work presented, attention was focused on:

- improvements in procedure when micropropagation is started from seed,

- attempts to induce micropropagation from phenotypically determined adult plants.





Material and methods

Buckwheat varieties »Darja« and »Siva« were used; seeds were harvested less than 1 year before sterilisation procedure. Plants from which parts were taken were grown under open field conditions.

Basal medium composition consisted of MS macro and micro elements (Murashige and Skoog 1962), 3 % sucrose, 0.6 % agar and (in mg/l): thiamine 2, pyridoxine 1, nicotinic acid 1, m-inositol 100, casein hydrolysate 2000. PH was adjusted before autoclaving to 5.7. Media were autoclaved for 15 min. at 121 °C and 15 PSI. Cultures were maintained under fluorescent lights with a day length of 16 hours.

Results

A. Procedures started from seeds

Since micropropagation is a final step also for other in vitro procedures some experiments were designed to optimise micropropagation stages.

It has been suggested earlier (Bohanec 1986) that when micropropagation is started from dehulled seeds, radicula should be removed after a few days and the rest of the plantlet should be placed on the medium to ensure close contact between the medium and shoot apex and so stimulate faster axillary branching. Data from experiment 1 (buckwhet variety »Darja«) show the differences in axillary bud formation in the treatments when radicula is not (A) and when it is (B) removed.

Experiment 1: Basic data and timetable

Basal medium, BAP 2 mg/l, IAA 0.2 mg/l

	procedure A	procedure B
No. of seeds incoluated	30	30
No. of ungerminated, albino or contaminated samples	3	4

In procedure B, radicula were removed 6 days after inoculation and the remaining plants placed on fresh medium with the same composition. Axillary buds were removed and weighted after 30 days from first innoculation in procedure A and after 16 days in procedure B. Fresh weight of axillary buds grouped in classes of 0.015 g is presented in Figure 1.

It can be seen that buds in procedure B have grown much faster and more uniformly. It was noticed that the apex grew coincidentally in a position close to the medium for the three more developed samples in procedure A. The induction phase was successful for almost all seeds grown by procedure B.

The media and procedures for subculturing axillary buds have been tested. It was observed that on media with unchanged cytokinin concentration as used for induction phase (BAP 2 mg/l, IAA 0.5 mg/l) more basal callus is formed than on media with lower citokinine concentration (BAP 1 mg/l, IAA 0.5 mg/l).

The propagation rate from dissected single shoots (2-3 cm long) placed on basal medium supplemented with BAP 2 mg/l and IAA 0.5 mg/l was analysed after 28 days of subculturing. Data of 30 shoots subcultured showed relatively large differences between formation of new, mostly axillary branches. New shoots have been counted separately as larger (over 2 cm) and smaller, on samples ready for placement on the rooting medium. An average $(\pm \delta)$ of 3.96 (± 2.33) larger and 1.43 (± 2.08) smaller buds have been formed from a single shoot. As seen from standard deviation, variability was very big in both classes. In 9 samples, flower buds started to form.

Instead of subculturing, a method of adding liquid medium following axillary bud formation (as in exp. 1 procedure B) could be a further simplification of the micropropagation procedure. In an experiment started with 30 seeds – variety »Darja« (9 ungerminated or contaminated) – procedure B described in exp. 1 was followed on media with the same composition. 6 days after inoculation, the radicula was cut and plantlets subcultured. 28 days after first inoculation, liquid medium (with the same composition but without agar) was added. 37 days after, it was estimated that the medium was exhausted. The result was an average of 8.91 (\pm 9.03) larger and 5.76 (\pm 7.99) smaller shoots, which developed from single seeds in 65 days. As seen from figures variability was very big also in these samples.

B. Procedures started from parts of adult plants

It was estimated that for plant breeding purposes, it would be of much greater value if it were possible to induce micropropagation from parts of plants at the time when they already have at least some ripened kernels, which is usually the moment at which the breeding value of individual plants could be observed.

14

At the time of ripening, buckwheat plants have only a limited number of green organs from which micropropagation could be started and even when they exist, it has been found very difficult to sterilise them. An attempt to use stem and leaf petiole cuttings failed because 99 % of inoculants have been contaminated.

A method of inducing new shoots from ripening plants was tested. Plants were cut to about 10 cm above ground, plants were transplanted to pots and well wetted. In 50 % of such plants new shoots and inflorescences appeared.

It had been estimated earlier that inflorescences would be the best organ for in vitro bud induction in buckwheat.

Several concentrations of NaOCl as sterilising agent were tested. The buckwheat variety was »Siva«. A treatment of 10 min. in 5 % NaOCl (with a few drops of surfactant Tween 20) followed by 3 rinses in sterile water was found satisfactory, since about 50 % of inocculums were not contaminated and about 30 % of those produced adventive or axillary shoots. In this case, the shoot induction medium consisted of basal medium supplemented with BAP 2 mg/l, KIN 2 mg/l and IAA 0.5 mg/l. The following micro-propagation stages were the same as when started from seeds.

Discussion

Since micropropagation is the basic tissue culture method, the experiments presented could help in optimising conditions for in vitro growth of buckwheat.

The preserving of valuable genotypes with known morphological characteristics by successful inoculation could enable transfer of cloned plantlets to field for controlled pollination, which could be the first use of buckwheat tissue cultures for breeding purposes. Certainly a real advance in stabilising plant characteristics can be expected only from dihaploid plants, which is the current research in some laboratories, as has been reported (Adachi, 1987).

Adachi, T. (1987): On the utility of plant tissue and cell culture in relation to plant breeding. Third Congress of Yugoslav Genetics with International Participation. Edt. N. Canki, Ljubljana, University School of Medicine, p. 60 (Abstract).

Bohanec, B. (1986): Present state and prospects of tissue cultures in breeding buckwheat. Buckwheat Research 1986, Proceedings of the Third International Symposium on Buckwheat, Pulawy, Poland, 7–12 July 1986, p. 45–56.

Murashige, T., Skoog, F. (1962): A revised medium for rapid growth and bio assays with tobacco tissue cultures. Physiol. Plant., 15, p. 473-497.

Nešković, M., Srejović, V., Vujičić, R. (1986): Buckwheat (Fagopyrum esculentum Moench.). Biotechnology in Agriculture and Forestry 2- Crops I. Edt. Y.P.S. Bajaj, Springer-Verlag, Berlin, Heidelberg, New York, Tokyo, p. 579-592.

Yamane, Y. (1974): Induced differentiation of buckwheat plants from subcultured calluses in vitro. Japan J. Genetics, 49, P. 139–146.

Izvleček

Izboljšave postopkov za mikropagacijo ajde (Fagopyrum esculentum Moench)

Za optimizacijo metode mikropropagacije ajde začete iz semena je bila preizkušena metoda odstanitve radikule 67 dni po *in vitro* kalitvi, čemur je sledilo uspešno aksilarno razraščanje, preizkušana je bila hitrost razraščanja aksilarnih poganjkov začetih iz posameznih poganjkov ter nadomestitev subkultiviranja z dolivanjem tekočga medija. Vpeljana je bila metoda temelječa na vzbuditvi rasti mladih poganjkov iz odrezanih starejših rastlin; iz novo nastalih socvetij je uspelo inducirati in vitro mikropropagacijo.

Light and temperature dependence of net photosynthesis for buckwheat

Lučka Kajfež-Rogataj

VTOZD za agronomijo, Biotehniška fakulteta, Univerza Edvarda Kardelja v Ljubljani, Ljubljana, Jugoslavija¹

Abstract

The dependence of net photosynthesis on light and on temperature was measured and analysed for the diploid cv. »Siva« variety of buckwheat.

Introduction

Many external factors influence CO_2 exchange between plant and environment. As a photochemical process net photosynthesis is directly dependent upon the availability of radiation. On the other hand the dark reactions of photosynthesis and respiration are biochemical processes limited especially by temperature. In spite of great importance of determining the influence of light and temperature on net photosynthesis for buckwheat, there are only a few works dealing with this problem (Akberdina 1970, Akberdina 1971, Populidi 1976, Kajfež-Bogataj and Gaberščik 1986).

Material

We estimated the effects of light and temperature on CO_2 exchange for buckwheat variety diploid cv. »Siva«. Plants were sown on 15 July and grown outside for 50 days. Two days before measurement they were put in the climate chamber in phenological stage of fully blossom. Regarding the prevailed weather, plants were adapted to rather high daily light intensities (daily sum of global radiation around 5 kWh/m²)· and to mean daily temperatures around 170°C.



Fig. 1: Light dependence of net photosynthesis in cv. »siva« (20°C and relative humidity 70 %).

¹ Agronomy Department, Biotehnical faculty. University E. Kardelj, Ljubljana, Yugoslavia.

Methods

Dependence of net photosynthesis upon light intensity was measured using Infrared gas analyser (IRGA, Type 225/2, Hoddeson, England). Artificial PAR radiation was simulated by Sylvania. GROLUX F14 TB, Denvers, USA and additional 1500 W lamp – TEZ Tesla, Yugoslavia) and measured by Quantum-Radiometer-Photometer (LI-COR, LI-185, Lambda, Nebraska, USA). Temperature in climate chamber was kept constant 20°C, which is very close to optimal temperature for net photosynthesis. Relative humidity was kept at 70 % and we used air with 350 to 400 ppm CO₂. First we measured leaf respiration in the dark and latter net photosynthetical activity at different light intensities. We raised (approximately doubled) the light intensity in time interval of half an hour from 200, 400, . . . to 40 000 lux.

The next step was determining the net photosynthesis relation to temperature. Leaf temperatures were measured by teletermometers (YSI, Model 44 TZ, Yellow Springs, USA). Artificial PAR radiation was simulated and kept constant at 25 000 lux (475 μ Em⁻²s⁻¹). Relative humidity was from 60 to 90 %. We raised temperature at the rate 5°C per hour. The low temperature and the high temperature interval were examined separately within period of 24 hours.

Results and discussion

The light dependence of net photosynthesis at optimal temperature (20°C) is given on Fig. 1. The compensation light intensity for buckwheat is rather low, bellow 500 lux (10 μ Em⁻²s⁻¹). Once the compensation point has been passed, CO₂ uptake increases rapidly till the light saturation point is reached between 35000 and 40000 lux. We can se that buckwheat net photosynthesis is not very efficient and thus the light dependence curve saturates at rather low intensity, being typical for C₃ plants. Maximum rate of CO₂ uptake at optimal temperature and saturated light intensity was close to 11 mg CO₂ dm⁻² hour⁻¹. Assuming the plants were healthy and in optimal stage of development regarding net photosynthesis capacity mentioned rate should be close to maximal CO₂ exchange capacity for cv. »Siva« variety. This study corresponds to results reported by Akberdina (1971).

The temperature dependence of net gas exchange (Fig. 2) shows three cardinal points: temperature



Fig. 2: The temperature dependence of net photosynthesis for cv. »siva« (light intensity 25 000 lux, relative humidity 70 %).

minimum for positive net, photosynthesis lies slightly bellow $O^{\circ}C$, which is relatively very low temperature in comparison with some other C₃ grain crops. Optimal range of temperature in which net photosynthesis is more than 90 % of maximum obtainable lies between 17° and 24°C. This interval is a bit narrower than in previous investigations (Kajfež-Bogataj and Gaberščik 1986). Such differences are normal and depend on ecological conditions of the plant (Larcher 1980). The high temperature limit for net photosynthesis or the heat compensation point is reached at 45°C. We can see that positive net gas exchange is measured over the wide range of temperatures which shows that cv. »Siva« variety is very well adapted to different thermal conditions which can appear in the environment.

References

Akberdina P.H., (1970): Vlijanje temperaturi vozduha na dnevnoj hod intenzivnosti fotosinteza i intenzivnosti eksomoza vodi iz listev grečihi. – Selekcija, genetika i biologija grečihi. Orel, 1970, 65–72.

Akberdina P.H., (1971): Transpiracijski koeficient fotosinteza u grečihi – Selekcija, genetika i biologija grečihi. Orel 1971, 182–187.

Kajfež-Bogataj L. and Gaberščik A., (1986): Analysis of net photosynthesis response curves for buckwheat. - Fagopyrum, Vol. 6, 1986, 6-8.

Larcher W., (1980): Physiological Plant Ecology, Springer Verlag, Berlin, 303 p.

Populidi K.H., (1976): Klimat i poživnjie posevi grečihi, Gidrometeoizdat, Leningrad, 1976, 114 p.

Izvleček

Ugotavljali smo odvisnost neto fotosinteze od svetlobe in temperature za diploidno ajdo cv. »siva«, v starosti 50 dni v fazi razvoja plodov. Svetlobna kompenzacijska točka za ajdo je pod 500 lux, saturacijska pa med 35000 in 40000 lux, merjeno pri optimalni temperaturi 20°C, minimalna temperatura za pozitivno neto fotosintezo je malo pod o°C, optimalni interval med 17° in 24°C, ter maksimalna temperatura pri 45°C.

Studies on buckwheat improvement¹

Prakash Narain

National Botanical Research Institute, Lucknow.

Buckwheat, besides a subsidiary food grain crop has also come into prominence as an exceptional plant containing protein of very high biological value. Buckwheat proteins are rich in many amino-acids as compared to other plant proteins, and are almost equal to animal proteins. In country like India, where people are largely vegetarial, buckwheat offers a great potential.

At present, buckwheat does not form part of diet of the majority of population and its use is restricted only to the people inhabiting the valleys in Himalayas where it is widely grown as a summer crop.

In the upper gangetic plains, it is also consumed in a variety of preparations especially on religious occasions after observing holy fast. It is still not grown as a regular crop. This is primarly due to the unsuitable climatic conditions in the plains for the crop and unremunerative returns. There is also lack of adequate planting material, particularly of the improved lines.

Considering the potential of the crop, a programme of work on buckwheat was undertaken at NBRI, Lucknow. The objectives were:

(I) Developping agro-techniques for buckwheat cultivation and

(II) Selection and evolution of new genotypes suitable to north Indian Plain.

Cultural Studies

The cultivation of buckwheat is restricted and mostly confined to the cool moist and temperate regions in the interior of Himalayas, ranging up to altitudes of 2000–12000 ft. However, preliminary observations have indicated that though buckwheat requires a moist and cold climate, it can also be successfully grown as a rabi season annual on the dry but well drained sandy and sandy loam soils of the northern plains, where day temperature does not exceed above 20°C from 15th December to 30th January. Buckwheat can be sown between 1st November to 15th November, and harvested in February.

The studies were undertaken for spacings and application of fertilizers. Plants were spaced 25-30 cm apart between rows and 10-15 cm from plant to plant. Seeding rate was 60 kg/ha. Fertilization was given at the rate of N 60 kg/ha, P₂O₅, 50 kg/ha, and K₂O 80 kg/ha and a total of 4 irrigations during the vegetative development period of 75 days were given. Observation were recorded on the effect of fertilizers on the height of plant, no. of flower cluster/plant and yield per 100 plants.

Application of urea resulted in an increase in height (4.03 %); no. of flower clusters (21.9 %) and in the yield of seed (5.8 %). Plants treated with superphosphate, reduced the height of plants (5.94 %) and seed weight (6.0 %) but increased (14.05 %) the no. of flower clusters. Muriate of potash had no significant effects either on height of plant and no. of branching or yield of grains.

Combined fertilizes gave better results than the ones, treated with urea, P_2O_5 and H K₂O, individually. There was an increase 3 %, in height of plants, 25.2 % in no. of flower clusters and 3.8 % in yield of grains.

Genetic improvement

A collection of 15 lines representing 6 varieties of buckwheat has been built up. Most of plants are acclimatized and are growing well.

Cytological analysis of different lines of *F. esculentum* indicated that all of them were diploids. In order to induce genetic diversity, some experiments to raise polyploids and induce mutations were also performed.

Polyploidy

In order to increase seed size and protein content, colchiploidy in three buckwheats was attempted and was successful. All the autotetraploids were cytologically studied. Meiotic analysis has indicated that they have irregular meiotic behaviour. Chromosome configurations like multivalents, bivalents and univalents have been observed occuring in variable frequencies at meta. I. The subsequent course of meiosis was also found irregular. Due to disturbances of genetic balance. There was reduction of pollen/ovule fertility and hence the seed setting in the colchiploids was low.

The C_3 generation of buckwheat polyploids are under study.

Studies to work out the co-relation ship between chromosome configurations chiasmata frequencies and seed set in the induced autotetraploid lines, are in progress.

¹ Presented at 3rd workshop of All India Coordinated Project on underutilized and unexploited plants, held at NBPGR, New Delhi, June 30 and July 1, 1986.

Mutations

Through X-ray irradiation, large number of phenotypic variations have been recorded within the mutation progenies. They are being assessed for their cytogenetical characteristics.

So far, 6 desirable lines with regards to their adaptability to local conditions, yield and quality characters, have been made. Their performance are under study.

Izvleček

V Indiji, kjer se ljudje prehranjujejo predvsem vegetarijansko, je ajda s svojimi kakovostnimi beljakovinami zelo pomembna potencialna poljščina. Za sedaj jo pridelujejo predvsem kot poletno poljščino v dolinah Himalaje. Na zgornjih gangeških planotah jo tudi uporabljajo v povezavi s postom. Na planotah jo lahko pridelujejo v zimskem času; sejejo jo v začetku decembra in žanjejo februarja. Opisani so poskusi pridelovanja in žlahtnjenja ajde (poliploidija, izzvane mutacije).

Aphelinus kashmiricus Hyat – a parasite of Aphis gossypii Glover (Melon Aphid) in Kashmir

Mohd. Rafiq Bhat

University of Kashmir, Srinagar-190006, India

Four species of buckwheats viz. Fagopyrum esculentum, F. tataricum, F. sagittatum and F. kashmirianum are cultivated in some remote and high altitude areas of Kashmir mainly because of nutritive value.

In Kashmir melon aphid-Aphis gossypii Glover (Homoptera: Aphididae) happens to be a serious pest of buckwheats causing substantial damage to plant through sucking sap. The infested leaves show curling, become wilted and finally dry up. Aphelinus kashmiricus Hyat (Hymenoptera: Aphelinidae) an endoparasite was reared from the adults of Aphis gossypii. In the laboratory parasites were obtained from field collected melon aphids infesting buckwheat plants. Bagat (1985) has recorded Aphelinus kashmiricus from Aphis gossypii attacking Zennia elegans in Kashmir.

Reference

Bagat, R.C. 1985, Geobios new Reports 4 : 178-179.

Study on the development and contributions of different plant organs to buckwheat dry matter production

Lučka Kajfež-Bogataj

VTOZD za agronomijo, Biotehniška fakulteta, Univerza Edvarda Kardelja v Ljubljani, Ljubljana, Jugoslavija*

Abstract

The field experiment was accomplished at Ljubljana in 1985 to study development and dry matter contributions of buckwheat roots, leaves, stems, flowers and grains to total dry matter production for cv. »Darja« during the growing period (18/07 - 07/10) through 7 successive samplings. Changes in dry weight of buckwheat plant organs and their relative contributions during the growth period are analysed and discussed.

Introduction

The growth rate of plant and of its organs during its development is variable. The increase in plant biomass is not the same in different phases of its growth. Knowledge of development characteristics of buckwheat plant is still incomplete in spite of several studies (Pauševa 1970, Ali-Khan 1973, Tsuzuki et al. 1983, Kajfež-Bogataj and Knavs 1985, Farooq and Tahir 1987). In order to reveal a more complete picture of the growth of different plant organs various growth analysis techniques can be applied (de Vries and van Laar 1982, Tsunoda 1983) based on field experiment. One of such approaches is presented in this work.



Fig. 1: Changes in dry weight of buckwheat plant organs during growth period.



Fig. 2: Relative contributions of buckwheat plant organs during growth period.

Material and methods

Diploid cv. »Darja«, plants were used in experiment. The buckwheat seeds were obtained from Agronomy Department, Biotehnical faculty, University E. K. Ljubljana. The plants were grown on the experimental field near Ljubljana from July 18 to October 7 in 1985. Fertilizer application rate was at 40 kg N, 40 kg P_2O_5 and 40 kg K_2O per hectar. Planting density was 200 plants per m². One plot was 60 cm \times 40 cm and consisted of two rows. The experiment was conducted in four replications.

During the growing period, sampling was carried out 7 times. The interval between sampling was 8-11 days (20, 31, 42, 50, 58, 66 and 76 days after germination, which was on July 24). Plants were collected together with roots and divided into roots, leaves, stems, flowers and grains. Dry weights were measured after drying 4 days at 60°C and 1 hour at 105°C before weighting.

* Agronomy Department, Biotehnical faculty, University E. Kardelj, Ljubljana, Yugoslavia.

Experimental results and discussion

The dry weights of different plant organs at various stages of growth development are presented in Table 1:

days after	dry weights (g/m ²)						
germination	leaf	stem	root	flowers	grain	total	
20	30.8	16.4	10.6	2.8	/	60.6	
31	40.7	67.2	15.6	13.8	/	137.3	
42	54.6	145.6	21.0	49.9	1	271.1	
50	57.8	187.0	26.2	71.3	43.8	386.1	
58	53.7	179.2	25.9	69.1	141.7	469.6	
76	28.2	183.7	27.7	73.2	179.2	492.2	

Table 1: Dry weights of different buckwheat organs in grams per m² ground

After germination the exponential growth phase began until 30th day when all the photosynthetic products were invested in the formation of new roots, leaves and stems. From 30th to 50th day growth rate remained nearly constant, the photosynthetic products being invested more in maintenance of the plant and less for expansion of for instance leaf area. After 50th day plant turned from the vegetative phase into the reproductive phase. The quantity of leaves is decreased, the quantity of stems and roots is stagnated but flowers and grains begin with further growth (Fig. 1).

During the development of a buckwheat plant, the partition of the photosynthetic product among its organs is important (Table 2). If organ growth is expressed as a relative contribution to the entire plant dry weight, a graph like that in Fig. 2 is obtained.

days after	relative contributions to total (%)							
germination	leaf	stem	root	flowers	grain	total		
20	50.7	27.0	17.7	4.6	/	100		
31	29.6	49.0	11.4	10.0	1	100		
42	20.1	53.7	7.8	18.4	1	100		
50	15.0	48.4	6.8	18.5	11.3	100		
58	11.4	38.2	5.5	14.7	30.2	100		
66	6.6	34.0	5.4	15.5	38.5	100		

Table 2: Relative contributions of different buckwheat organs in % of total dry weight

The relative shares of leaves and roots decrease exponently with time during the buckwheat development. The stems reach maximum percentage near 40th day after germination, later on their relative contribution decreases. The relative contribution of flowers increases till 50th day and later on slowly decreases while grains develop. The relative dry weight of grains to total dry weight show an exponental increase and in the end of growth period gives a crop index about 36 %.

Conclusions

In buckwheat the partitioning of assimilates to different organs and especially the partitioning of dry matter between grains and vegetative parts is of great importance. About the internal control of distribution of assimilates in buckwheat still little is known. This implies that more data about dry matter distribution have to be collected experimentally. Experimental data should give a certain key for dry matter distribution among plant organs. Such a key would be characteristic for studied variety, although it may be different for different cultivars.

Reference

Ali.Khan, T.S. (1973): Effect of row spacing and seeding rate on yield of buckwheat. Agron. Jour. vol. 65, 914-915.

Farooq, S. and Tahir, I. (1987): Comparative study of some growth attributes in buckwheats (Fagopyrum Spp.) cultivated in Kashmir, Fagopyrum 7., 11–14.

Kajfež-Bogataj, L. and Knavs, M. (1985): Studies on the production of dry matter in the community of buckwheat with particular reference to leaf area, Fagopyrum 5, 1985, 7-3.

Pauševa, Z. P. (1970): Osobenosti cvetenija i plodoobrazovanja u grečihi v svjazi s javleniem nesovmestimosti. Selekcija i agrotehnika grečihi, Orel 1970, 21–30.

Tsunoda, S. (1983): Designing the green to harvest the sun, Tohoku University, Sendai, Japan, 298p. Tsuzuki, E., Shimiya, M. and Shida, S. (1983): Studies on the dry matter production of buckwheat. Proc. 2nd Int. Symp. Buckwheat Myazaki, 165-172,

42 meteric contribution of a sector formation of a sector of a sector of the sector

de Vries, P. and van Laar, H. H. (1982): Simulation

of plant growth and crop production, Pudoc Wageningen, 308 p.

Izvleček

Opisani so rezultati, dobljeni na osnovi poljskega poskusa z genotipom »Darja« v Ljubljani v letu 1985 (Setev 18/07 – žetev 07/10). Z zaporednimi sedmimi odvzemi vzorcev smo ugotavljali rast in razvoj posameznih rastlinskih organov (korenin, listov, stebel, cvetov in zrnja). Prikazana sta poteka absolutnih in relativnih prispevkov mase posameznih organov k skupni masi ajde tekom rastne dobe ter opisane njune glavne značilnosti.



INTERNATIONAL BUCKWHEAT RESEARCH ASSOCIATION

Present seat: Institute of Soil Science and Plant Cultivation in Puławy - Poland

Adress: Instytut Uprawy, Nawożenia i Gleboznawstwa Osada Pałacowa, 24-100 Puławy telephon: 34-21 34-22 telex: 0642410

The information

about the General Assembly of IBRA members which took place during the 3rd International Buckwheat Symposium, Puławy 7–12 of July, 1986.

On the meeting discussed the rules of IBRA, activity of the Association in the future, edition of New sletter »Fagopyrum«, continuation of agroecological study, organization of next symposium and at the end of the meeting it was elected the new IBRA Executive Committee.

1. All members of Association agree with rules which were accepted on IBRA general assembly in Miyazaki, Japan, during the 2nd International Buckwheat Symposium in September 1983.

2. The activity will be developed mainly in the following fields:

a) Edition of New sletter Fagopyrum (Prof. Dr. I. Kreft and his coworkers). Publishing and editing is autonomous, but proposal and advice of IBRA executive committee are taken into account. The free copies of Fagopyrum are distributed to scientists and members of IBRA. »Fagopyrum« is very important newsletter because it is possible to present the newest information about various fields of buckwheat studies. The articles, announcements, informations can be in English or in other language with summary in English.

b) The agroecological studies will be continued in 1987 and 1988. Every year results will be send to the representatives of various countries.

c) The studies on the problems of buckwheat grain, groats and flour quality. Special group from Jugoslavia (Doc. Dr. B. Javornik), Poland (Doc. Dr. Ł. Fornal, Prof. Dr. M. Ploszyński) and Denmark (Dr. B. O. Eggum) will prepare the proposition for discussion. The proposition will be published in »Fa-gopyrum«. Remarks will be colleted by editorial staff of »Fagopyrum« and send to the authors. The »quality group« prepare the final conclusion and present on the 4th International Buckwheat Symposium and also in New sletter »Fagopyrum«. This group will discuss also the problems of new products prepared from the buckwheat (grains, plant, leaves, flowers, hulls).

d) Will be lead in Poland the collection of different species, varieties, cultivars or genetic lines of buckwheat. Everybody who is interested on that project can get the sample of the seeds. We ask the scientific workers who has interesting lines of buckwheat to send them to the head of the collection (Dr. Kalina Komenda, High School of Agriculture and Pedagogics, Department of Genetics, Prusa Street 12, 08-110 Siedlee, Poland). It will be also very useful to publish the information about the different new lines of buckwheat in New sletter »Fagopyrum«.

e) It is proposition to organize two groups connected with: storage of buckwheat seeds and protection the plants against diseases and insects. We are waiting for information who is especially interested in that project and one can be organizer of the scientific workers group which will prepare the scientific papers (lectures or announcements) on the 4th buckwheat symposium.

3. The next symposium on buckwheat will be organized by USSR (Professor Dr. N. V. Fesenko, All-Union Research Institute of Grains and Legumes, Orel, p/o Strieleckoje/ in July 1989. It is also proposition that the 5th International Buckwheat Symposium will be in India in 1992.

4. The General Assembly on the session during the 3rd International Buckwheat Symposium elected the chairman, vicechairmen and other members of IBRA Executive Committee for the years 1986–1989.

The Executive Committee

Chairman Vice-chairman Vice-chairman The members Prof. Dr. Marek Ruszkowski Prof. Dr. Michal Ploszyński Doc. Dr. Marian Król Prof. Dr. Krystyna Kusiorska Doc. Dr. Jan Mazurek Doc. Dr. Łucja Fornal Doc. Dr. Jan Szklarz Dr. Kalina Komenda

We are waiting for continuation of »buckwheat family« cooperation. Every problem connected with buckwheat genetics, physiology, breeding, technology of production, quality and utilization we ought to present in Newsletter »Fagopyrum« or send as a information to the IBRA Executive Committe in Poland (President of IBRA, Prof. Dr. Marek Ruszkowski, Institute of Soil Science and Plant Cultivation, Czartoryskich Street 4, 24-100 Pulawy, Poland).

For sure our Association will develop and the members will cooperate thorugh Newsletter »Fagopyrum«, Executive Committee and International Symposiums.

The best regards for all members of IBRA and whole »buckwheat family«. We are waiting for your cooperation.

Yours sincerely Marek Ruszkowski President of IBRA



BUCKWHEAT RESEARCH 1986

Proceedings

of the 3rd International Symposium on Buckwheat Pulawy, Poland, 7-12 July 1986

CONTENTS

Pai	1, 1, 1 and $1, 2, 2$ and $1, 2, 2$ and $1, 2, 2$ and $2, 2, 2$ and $2, 2, 2$ and $1, 2, 2, 3$ and $1, 2, 2, 3$	
		Page
1.	Król M. History of buckwheat cultivation in Poland	7
2.	Alekseeva E. S. Selection, cultivation and utilization of buckwheat	18
3.	Kreft I. Physiology of buckwheat yield	37
4.	Javornik B. Buckwheat in human diets	51
5.	Ruszkowski M. Productivity of buckwheat	78
6.	Fesenko N. Buckwheat breeding for stable high yielding	99
7.	Adachi T. Is it possible to overcome the low yield of buckwheat by means of biotechnology?	
	Some trials in plant genetic manipulation	106
8.	Kusano T., Chiu H., Ikeda K., Oku M., Physicochemical properties of buckwheat proteins	117
9.	Komaki M.K. Inbreeding depression and interaction among deleterious genes	
	in buckwheat populations	128
10.	Luthar Z., Kocjan D., Kreft I. Breeding buckwheat with determinant growth habit	139
11.	Taranenko Z. Heterosis in the buckwheat selection	145
12.	Dovzhenko L.I., Melkonova Ye., Kuzmina A.V. Possible application of cytogenetic	
	analysis in the selection of tetraploid buckwheat	154
13.	Kusiorska K., Adamkiewicz E., Samborska-Ciania A. Research works of a fructifing	
	process of Fagopyrum esculentum depending on the type of flower pollination	155
14.	Munshi A.H. The taxonomy of genus Fagopyrum / Polygonaceae /	
	and its species delineation in Himalayas	162
15.	Górski T. Buckwheat yield dependency on climatic conditions	169
16.	Namai H. Pollination biology and seed multiplication method of buckwheat genetic resources	\$ 180
17.	Ohnishi O. Linkage relationships among the mutant lines established in common buckwheat.	
	Fagopyrum esculentum Moench	187
18	Ohnishi O. Evaluation of world buckwheat varieties from the view points of genetics.	
	breeding and the origin of buckwheat cultivation	198

Part II.

1.	Ren Shuhue, Wang Zhunging Liu Anlin. Observation on flowering habits	
	of common buckwheat	5
2.	Ren-Shuhua, Liu Anlin. The survey of cultivated buckwheat, pollen spreading distance	
	and the relation between pollen carried by insects and yield	10
3.	Mukhiya Y. K., Singh V. P. Ascorbic acid oxidase activity in common buckwheat	
	in relation to manganese treatments	18
4.	Gohil R. N., Misri B. Comparative performance of eleven local buckwheat germplasms	
	under uniform environmental conditions	23
5.	Kajfež-Bogataj L., Hočevar A. Trials to model growth of buckwheat plant canopy	
	from dynamic point of view	29
6.	Bohanec B. Present state and prospects of tissue cultures in breeding buckwheat	45
7.	Skrzypczak L., Thiem B. Fagopyrum esculentum tissue culture and the control	
	of active compounds content	57

Page 8. Warcholowa M., Mroczkowski W., Nitrogen, potassium and magnesium nutrition and mineral composition of buckwheat 59 9. Komenda B., Komenda K., Komenda-Ronka J. Study of the influence of herbicides as successive buckwheat crops 66 10. Komenda K., Komenda-Ronka J., Komenda B. Study of buckwheat varieties collection 68 11. Kusiorska K., Szczukowski S., Tworkowski J. The effect of Betakson on buckwheat yield and seed sowing quality 71 12. Wang Zhunging Liu Anlin, Ren Shuhua. Study on the high-yialding cultivation of buckwheat. 79 13. Fatyga J. The influence of various technologies of buckwheat growing on the quantity 95 and quality of yield 14. Szklarz J., Olender K. Effect of different mineral fertilization on utilitarian features and seed yield of buckwheat 100 15. Ruszkowski M., Zebrovski Z. The productivity of homomorphic and heteromorphic buckwheat forms in various agrotechnical conditions 105 16. Ikeda K., Oku M., Kusano T., Chiue H. Antinutritional substance in buckwheat seeds. 110 17. Dunaevsky S. A., Belozersky M. Proteolysis of the buckwheat seed storage globulin 115 18. Mazza G. Factors influencing storage quality and dehulling characteristics of buckwheat 120 19. Bubicz M., Korzen A., Korzeń S. Effect of increasing nitrogen rates on the chemical composition of buckwheat seeds 121 20. Jablonski B., Szklanowska K., Ruszkowski M. Apiarian quality of homostyle buckwheat and effect of bees on its yield 126 21. Fornal L., Soral-Smietana M. Chemical composition and utilization of buckwheat in feeding technology 134

22. Bubicz M., Sklarz J. Effect of selected micronutrients on the growth, yield, and chemical composition of buckwheat
23. Dietrych-Szostak D., Ploszyński M. Chemical composition and feeding value of buckwheat hulls and harvest residues
149

24. Dietrych-Szóstak D., Ploszyński M. Some aspects of the nutritional value of light and dark buckwheat groats

Part III.

1. Welcome address of president of Pulawy City	5
2. The address of IBRA chairman	7
3. The address of the Organizing Committee	8
4. Welcome address: director of Institute of Soil Science and Plant Cultivation	12
5. Sokolov O. A. Mineral nutrition on buckwheat in ontogenesis	16
6. Hagami S. Mt. Hiei and buckwheat	22
7. Ruszkowska M. Aphids on buckwheat and possibility of their control	29
Supplement	
1. Research group for collective study on buckwheat in Poland	5
2. Peasant proverbs connected with buckwheat cultivation	7
3. Polish dishes from buckwheat	11

156

MAILING LIST – SEZNAM NASLOVOV

Professional interest of scientists is preliminary marked according the proposal published in Fagopyrum 6: p. 47 (1986). Any changes or additions should be noticed to the Editor.

Additional copies are distributed to students, at scientific symposia, to public libraries in Yugoslavia and abroad etc.

Dr. Taiji Adachi

ipt

i

0

d

Laboratory of Plant Breeding Faculty of Agriculture Miyazaki University, Kumano 7710 Miyazaki 889-21 JAPAN

Dr. France Adamič Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

Dr. E. S. Alekseeva nps Selskohozjajstvenij Institut Ul. Ševčenko 13, 281900 G. Kamenec-Podolskij, Hmelnickoj obl. SSSR

Dr. P. Apel

Akademie der Wissenschaften der DDR, Zentralinstitut für Genetik und Kulturpflanzenforschung 4325 Gatersleben DDR

Dr. Radik M. Avezdzhanov

Chief, Buckwheat section N.I. Vavilov V.I.R. Gercena, 44 190000 Leningrad SSSR

Dr. Momčilo Babić Apartado Postal 55 Catacamas HONDURAS, C.A.

Dr. Ramesh Kumar Bali Department of Zoology University of Kashmir SRINAGAR – 190006 INDIA **Dr. Zoltán Barabás** Cereal Research Institute 6701 SZEGED, POB. 391 MADŽARSKA

Dr. N. El Bassam Institut für Pflanzenbau und Pflanzenzüchtung der Bundesforschungsanstalt für Landwirtschaft (FAL) Bundesallee 50 D-3300 Braunschweig

BR DEUTSCHLAND

Dr. M. Belozersky A. N. Belozersky Laboratory of Molecular Biology and Bioorganic Chemistry, Moscow State University Moscow SSSR

Dr. Mohd. Ragiq Bhat Department of Zoology University of Kashmir SRINAGAR – 190006 INDIA

Bibliography of Agriculture National Agricultural Library, USDA U.S. Department of Agriculture Washington D.C. USA

Bibliotheek

Institut voor Plantenveredeling Postbus 386 6700 AJ WAGENINGEN NEDERLAND

Bibliothek Landbouwhogeschool P.O.Box 9100 6700 HA WAGENINGEN NEDERLAND d

r

Steen Blicher acirs Fagodan A/S Dalkildegaardsallé 1 DK-5600 Faaborg DANMARK

Dr. Miroslav Bogdanović ips

Poljoprivredni fakultet Sarajevo Zagrebačka 18 71000 Sarajevo JUGOSLAVIJA

Dr. N. A. Bogkarev Laboratorija po grečihi Selskohozjajstvenij Institut Ul. Ševčenko 13, 281900 G. Kamenec-Podolskij, Hmelnickoj obl. SSSR

Borut Bohanec Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

The British Library The Science Reference Library Holborn Branch 25 Southampton Buildings LONDON, WC2A 1AW GREAT BRITAIN

Dr. M. Bubicz Katedra Chemii Ogólnej i Biochemii Akademia Rolnicza Lublin, Lublin POLAND

Bulletin Signaletique Centre de Documentation du C.N.R.S. 15 quai Anatole-France 75-Paris-VIIe FRANCE

Dr. G. M. Butt Department of Botany University of Kashmir Srinagar - 190006 INDIA

CABS Current Awareness in **Biological Sciences** 132 New Walk, Leicester LE1 7QQ ENGLAND, G.B.

Dr. Clayton Campbell Agriculture Canada

Research Station, Box 3001 MORDEN, MANITOBA, ROG 1JO CANADA

Centre for Agricultural Publishing and Documentation - PUDOC 17 Marijkeweg, P.O.Box 4, 6700AA Wageningen THE NETHERLANDS

Dr. Hiroko Chiue Faculty of Nutrition Kobe-Gakuin University Ikawadani-cho, Nishi-ku, Kobe 673 JAPAN

Miloš Chladek Fučikova 14 776 Olomuc-6 ČSSR

C. P. Christensen

airs

cr

Fagodan A/S Dalkildegaardsallé 1 DK-5600 Faaborg DANMARK

Dr. F. Le Cochec Centre de Recherches de Rennes Station d'Amélioration des Plantes Domaine de la Motte-au-Vicomte 35650 LE RHEU – B.P. 29 FRANCE

Commonwealth Bureau of Pastures and Filed Cropss Hurley – Maidenhead Berks SL6 5LR - UK ENGLAND, G.B.

inpt

r

Commonwealth Bureau of Plant

Breeding and Genetics, Dept. of Applied Biology Pembroke Street, **Cambridge,** CB2 3DX ENGLAND, G.B.

Mieczyslaw Dabrowski

Wysza Szkola Rolniczo Pedagogiczna w Siedlcach ul. B. Prusa 12 08-110 Siedlce POLSKA

Dr. H. De Jong

Agriculture Canada Research Station P.O.Box 20280 FREDERICTON, N. B. E38 4Z7 CANADA

Mr. Knud Dencker-Jensen aci Staagerupvej 48 DK-5762 Vester Skerninge DANMARK

Dr. Natalia I. Dermileva

Selskohozjajstvenij Institut Ul. Ševčenko 13, 281900 G. Kamenec-Podolskij, Hmelnickoj obl. SSSR

Department of Plant Breeding

University of Helsinki 0071 Helsinki 71 SUOMI – FINLAND

Dokumentationsstelle für

pflanzliche Produktion D 8050 Freising-Weichenstephan BR DEUTSCHLAND

Dr. Dorota Dietrych-Szóstak abr Instytut Uprawy, Nawozenia i Gl. Osada Palacowa, 24-100 Pulawy POLAND

Dr. Slava Doberšek-Urbanc

Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

Dr. L. I. Dovzhenko

N. K. Koltzov Institute of Developmental Biology Akademija Nauk SSSR 26 Vavilov st., 117808 Moskva SSSR

Dr. Y. Dunaevsky

A. N. Belozersky Laboratory of Molecular Biology and Bioorganic Chemistry, Moscow State University Moscow SSSR

Dr. Hans Eggers

Inst. f. Landw. Botanik der Universität Meckenheimer Allee 176 D-5300 Bonn 1 BR DEUTSCHLAND

abr

ir

Dr. B. O. Eggum Statens Husdyrbrugsforsg Rolinghedsvej 25 DK – 1958 Kobenhavn V DENMARK

E.N.S.A.R. Bibliothéque Centrale 65, rue de Saint-Brieuc 35042 RENNES CEDEX FRANCE

FAEP – Federacao da Agricultura do Estado do Paraná
DEES, Marechal Deodoro, 450-14°, C.P. 2744
CEP 80.000-CURITIBA-PARANÁ
BRAZIL

FAO Library Viale Terme di Caracala Roma ITALIA ip

Dr. Sikandar Farooq Department of Botany University of Kashmir Naseembagh SRINAGAR 190006 INDIA

, du

1,2,4n

ir

r

krs

Dr. Józef Fatyga Akademia Rolnicza we Wroclaviu 50-373 Wroclaw POLAND

Dr. A. C. Ferault defgh INRA Route de St. Cyr 78000 VERSAILLES FRANCE

Dr. N. V. Fesenko np Vsesojusnij naučno-issledovatelskij institut zernobobovih i krupjanih kultur Orel 303112 SSSR

Filial Biblioteki Akademii Nauk SSSR Baltiskaja ul. 14 Moskva A-219 SSSR

Nina Flašar Bul. revolucije 249 11000 Beograd

JUGOSLAVIJA Dr. Łucja Fornal

Akademia Rolniczo-Techniczna w Olsztynie Zaklad Technologii Produktow Pochodzenia Roslinnego 10-957 Olsztyn 5 POLSKA

Dr. Alicia de Francisco Carlsberg Laboratory Gamle Carlsberg Vej 10 DK-2500 Kobenhavn Valby DANMARK Alenka Gaberščik Inštitut za biologijo Univerza E. Kardelja, Ljubljana Aškerčeva 12, 61001 Ljubljana

Aškerčeva 12, 61001 Ljubljana JUGOSLAVIJA

Dr. R. N. Gohil Cytogenetics Laboratory Department of Botany University of Kashmir SRINAGAR-190 006 INDIA

Dr. Evgenya Gorina Belorusskij naučno-issledovateljskij institut zemledelija BSSR 222160, g. Jodino Minoski obl. SSSR

Dr. Tadeusz Górski

Instytut Uprawy, Nawozenia i Gl. . Osada Palacowa, 24-100 Pulawy POLAND

Dr. Daniela K. Gotsova Laboratory of Poliploidy Institute of Genetics 1113 Sofia, P.O.Box 96 BULGARIA

Dr. Mario Gregorič Kmečka zveza – Trst Ul. Cicerone, 8/B Trieste ITALIA

Dr. G. H. Gubbels Research Station, Agriculture Canada **Morden,** Manitoba ROG 1JO CANADA

Dr. Marek Grzywa IHAR Ul. Zawila 4 KRAKOW – 12 POLSKA 124

k

i

Dr. Mohamoud Hamoui University of Aleppo Research Center of Aleppo University of Agriculture ALEPPO SYRIA

Dr. P. van Heeuwen ZWAAN & de WILJES B.V. Stationsstraat 124 9679 ZG SCHEEMDA (GR) PAYS – BAS, HOLLAND

Todaki Hirose Depertment of Botany Faculty of Science University of Tokyo Hongo Tokyo 113 JAPAN

Dr. Andrej Hočevar Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

Dr. Park Cheol Ho Department of Agronomy Kangweon National University Chuncheon, 200 KOREA (South Korea)

Hortus centralis Facultas Medica Universitas Purkyniana, Komenského nám. 2 66243 Brno ČSSR

ICARDA P.O.Box 5466 Aleppo SYRIA

INRA – Service de Documentation Centre de Recherches Agronomiques Route de St. Cyr, 78000 VERSAILLES FRANCE

K. Irie

Laboratory of Plant Bréeding Faculty of Agriculture Miyazaki University Funatsuka 3-210, Miyazaki 880 JAPAN

Kiyokazu Ikeda Faculty of Nutrition Kobe-Gakuin University Tarumi Kobe 673 JAPAN

Institute for Agricultural Research, Samaru Ahmadu Bello University P.M.B. 1044, ZARIA NIGERIA

Institut für Lebensmittelwiss Eidg Technische Hochschule Universitätstrasse 2 8006 Zürich SCHWEIZ

k

Dr. Branka Javornik Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

Dr. Freeman K. Johnson Johnson Foundation Seed 731 Homestead Avenue Moorhead, Minnesota 565560 USA

acrt

ps

i

Ms. Inge Jorgensen Statens Planteavls Forsogstationen Ledreborgallé 100, DK-4000 Roskilde DENMARK

Dr. Marijan Jošt Poljoprivredni inštitut Križevci 43260 Križevci JUGOSLAVIJA

ar

р

i

Dr. inž. Roman Jurga

0051196 Centralne Laborator. Technologii Prezetworstwa i Prezechowalnictwa Zbož 00-041 Warszawa ul. Jasna 14 POLSKA

Lučka Kajfež

Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

A. Kajimoto

Laboratory of Plant Breeding Faculty of Agriculture Miyazaki University Funatsuka 3-210, Miyazaki 880 JAPAN

Dr. S. J. Kalembasa Agricultural and Teachers University, ul. 3. maja 54 08-110 Siedlce POLSKA

Dr. N. Katayama Laboratory of Genetics Faculty of Agriculture Kyoto University Kyoto 606 JAPAN

Dr. K. Kawabata Laboratory of Plant Breeding Faculty of Agriculture Miyazaki University Funatsuka 3-210, Miyazaki 880 JAPAN

Dr. Keiji Kenjo Dept. of Land Development College of Agriculture and Veterinary Medicine, Nihon University 3-34-1, Shimouma Setagayaku JAPAN ar

k

p

n

f

Knjižnica SAZU Novi trg 3 61000 Ljubljana JUGOSLAVIJA

Darja Kocjan

Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

Kyoko Koki

1-39, Otsubo Nishi 1-Chome Miyazaki City, Miyazaki, 880 JAPAN

Dr. Boguslaw Komenda Stacja Hodowli Rošlin Jeleniec 21-422 Stanin POLSKA

Joanna Komenda-Ronka

Wysza Szkola Rolniczo-Pedagogiczna w Siedlcach ul. B. Prusa 12 08–110 Siedlce POLSKA

Dr. Kalina Komenda

Instytut Biologii Stosowanej Wysza Szkola Rolniczo-Pedagogiczna ul. B. Prusa 12 P.08-110 Siedlce POLSKA

Dr. Svetlane K. Kirillenko

Sel'skohozjajstvennij institut ul. Sevčenko 13 g. Kamenec-Podol'skij Hmel'nickoj oblasti, 281900 SSSR

A. Korzeń

Katedra Chemii Ogólnej i Biochemii Akademia Rolnicza Lublin, Lublin POLAND

iop

iop

f

S. Korzeń

Katedra Chemii Ogólnej i Biochemii Akademia Rolnicza Lublin, Lublin POLAND

Mieczyslaw Koter

Instytut Chemizacji Rolnictwa Akademii Rolniczo-Technicznej w Olsztynie Olsztynie POLSKA

Viktor I. Kovalenko

Institute of Cytology and Genetics Academy of Sciences of the USSR Siberian Division 630090 Novosibirsk SSSR

Dr. Teresa Koszykowska

Institute of Plant Breeding Agricultural and Technical Academy Olsztyn-Kortowo POLSKA

Ivan Kreft

Biotehniška fakulteta Univerza E. Kardelja Krekov trg 1 61000 Ljubljana JUGOSLAVIJA

Marian Król

Instytut Uprawy, Nawozenia i Gl. Osada Palacowa, 24-100 Pulawy POLAND

Dr. Roman Kubiczek Ogrod botaniczny PAN ul. Prawdziwka 2 P-00-979 Warszawa p-84 POLSKA

Dr. Krystyna Kusiorska

Institute Of Plant Breeding Agricultural and Technical Academy Olsztyn-Kortowo POLSKA

r

ir

p

ainpr

0

p

Dr. Takanori Kusano Faculty of Nutrition Kobe-Gakuin University

Tarumi Kobe 673 JAPAN

Laboratory of Plant Breeding Faculty of Agriculture Miyazaki University, Kumano 7710 Miyazaki 889-21 JAPAN

Dr. Siefgried Lachmann

Antonia-Viscontistr. 24 D-7120 Bietigheim-Bissingen BR DEUTSCHLAND

Library

Carlsberg Laboratory Gamle Carlsberg Vej 10 DK-2500 Kobenhavn Valby DANMARK

Library Indian Agricultural Research Institute NEW DELHI – 110012 INDIA

Library

Kobe-Gakuin University Ikawadani-cho, Nishi-ku, Kobe 673 JAPAN

Dr. Anlin Liu The Agriculture Academy of Science of Inner Mongolia Huhehaote City, Inner Mongolia P.R. CHINA

Zlata Luthar Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA inp

ar

defgh

Laboratory of Plant Breeding Dr. James Mac Key Department of Genetics and Plant Breeding Agricultural University of Sweden S-75007 UPPSALA

SWEDEN

Dr. Eizo Maeda Faculty of Agriculture Nagoja University, Furocho, Chkusa, Nagoya, 464 JAPAN

Dr. G. Mazza Agriculture Canada, Research Sta. Morden, Manitoba, ROG IJO CANADA

Dr. Juju Bhai Manadhar defgi Division of Plant Pathology Khumaltar, Lalitpur NEPAL

Dr. Andrej Martinčič Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

Kazimierz Majkowski Instytut Uprawy Roli i Roslin AR-T w Olsztynie Zaklad Roslin Zbozowych i Przemyslowych Polsztyn-Kortowo POLSKA

Dr. Toshiko Matano aiop Department of Crop Science and Plant Breeding Faculty of Agriculture Shinshu University Ina, Nagano-Ken JAPAN

Dr. Tetsujiro Matsuhashi ar Nagano State Laboratory of Food Technology, Kurita, Nagano-shi, 380 JAPAN Dr. T. M. Mizukubo

pr

rs

k

Laboratori of Plant Breeding Faculty of Agriculture Miyazaki University Funatsuka 3-210, Miyazaki 880 JAPAN

Haruyo Miyashita Faculty of Nutrition Kobe-Gakuin University Tarumi-ku Kobe JAPAN

Toshitaka Miyazaki Laboratory of Erosion Control Engineering Faculti of Agriculture Shinshu University JAPAN

Dr. Lea Milevoj Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

Dr. Miguel Motta Agricultural Institute OEIRAS PORTUGAL

Dr. A. H. Munshi Department of Botany University of Kashmir Srinagar – 190006 Kashmir INDIA

Dr. Y. K. Mukhiya School opf Studies in Botany Vikram University Ujjain 456010 INDIA

Dr. Lars Munck Carlsberg Laboratory Gamle Carlsberg Vej 10 DK-2500 Kobenhavn Valby DANMARK ar

Yuichi Nagata Shibata Publishing Company 3-33-5 Hongo, Bunkyo-ku Tokyo JAPAN

Dr. Takashi Nagatomo 7075 Funahiki Kiyotake-cho Miyazaki, 889-16 JAPAN

Kazuko Nakano Faculty of Nutrition Kobe-Gakuin University Tarumi-ku Kobe JAPAN

Dr. Hyoji Namai ainopqs Institute of Agriculture and Forestry, University Tsukuba Sakura, Niihari, Ibaraki 305 JAPAN

Dr. Prakash Narain Cytogenetics Laboratory National Botanical Research Institute, LUCKNOW-226001 INDIA

Dr. Mirjana Nešković Biološki institut S. Stanković Ulica 29. Novembra 11000 Beograd JUGOSLAVIJA

Dr. Kiyoshi Nishimaki Chushin Agricultural Experimental Station Nagano Agricultural Research Centre 399-07 Shiojiri-shi Nagano-ken JAPAN

Dr. Kazimierz Noworolnik IUNG, ul. Czartoryskich 4 Pulawy POLSKA ainpt

ar

ainp

kt

Dr. Ohmi Ohnishi Laboratory of Genetics Faculty

ainop

ir

ar

Laboratory of Genetics Facult of Agriculture Kyoto University Kyoto 606 JAPAN

Tomotada Ono Department of Agricultural Chemistry Iwate University MORIOKA JAPAN Dr. Miho Oku Faculty of Nutrition Kobe-Gakuin University Ikawadani-cho, Nishi-ku, Kobe 673 JAPAN

Dr. Krystyna Olender Akademia Rolnicza w Lublinie Lublin POLAND

Jakob Opperer jun. Thalmann 5 D-8201 Rohrdorf BR DEUTSCHLAND

Dr. Zoja Petrovna Pauševa Ul. Kosmonavtov, 4, kv. 167 129 366 Moskva SSSR

Dr. Birthe Pedersen Carlsberg Laboratory Gamle Carlsberg Vej 10 DK-2500 Kobenhavn Valby DANMARK

Dr. Michal Ploszyński Instytut Uprawy, Nawozenia i Gl. Osada Palacowa, 24-100 Pulawy POLAND ar

abr

PNS – UED Pošta 022 Bratislava ČSSR

Ksenija Pundrić inop Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

Dr. K. P. Narasimha Rao 795 »Krishna Kunj«, 36 Cr Cross 18th Main, Tayanagar, IV. T. Block Bangalore-11 INDIA

Dr. Ngamchuen Ratanadilok

Faculty of Agriculture Kasetart University Bangkhaen Campus Bangkok 10900 THAILAND

Dr. R. Reimann-Philipp Bundesforschungsanstalt für gartenbauliche Pflanzenzüchtung Bornkampsweg 31 2070 AHRENSBURG BR DEUTSCHLAND

Matti Rekunen

Hankkijan Kasvinjalostuslaitos SF-04300 HYRYLÄ SUOMI – FINLAND

Dr. Shuhua Ren io The Agriculture Academy of Science of Inner Mongolia Huhehaote City, Inner Mongolia P.R. CHINA

Dr. Robert G. Robinson University of Minnesota Department of Agronomy and Plant Genetics Agronomy Building 1509 Gortner Avenue St. Paul, Minnesota 55108 USA Dr. Barbara Ruszkowska ainp Zaklad Roslin Zbozowych IUNG Pulawy POLSKA

Dr. Marek Ruszkowski ainop Zaklad Roslin Zbozowych IUNG Pulawy POLSKA

Dr. Rubens S. F. do Amaral R. Guilherme Guimarães, 212 80 000 – CURITIBA – PARANÁ BRASIL

Dr. B. C. Saha Agric. College P.O. Sabour, BHAGALPUR BIHAR-813210 INDIA

Dr. Takahiko Sakagami Iriya 1-30-7, Taito-ku, Tokyo 110

JAPAN

op

inp

ar

C

Sandra Salmins 9A Lamarche Ste Anne de Bellevue, P.Q. H9X 2A1 CANADA

Danuta Setlak Wysza Szkola Rolniczo-Pedagogiczna w Siedlcach ul. B. Prusa 12 08-110 Siedlce POLSKA

Riichiro Shiratori

7-9-35, Tsudanuma Narashino-City Chiba-Prefecture JAPAN

Dr. V. P. Singh School of Studies in Botany Vikram University Ujjain 456010 INDIA ar

kr

Dr. Lutoslawa Skrzypczak ct Katedra Botaniki Farmaceutycznej Akademia Medyczna im. K. Marcinkowskiego, Wieniawskiego 1 61-712 Poznań POLAND

38

Dr. O. A. Sokolov Institut agrohimii i počvovodenja Puščino Moskv. obl. 142292 SSSR

Mme Marion Sorin Exchange Section INRA, Institute National de la Recherche Agronomique Unité Centrale de Documentation R. St. Cyr-F 78000 Versailles FRANCE

Dr. Jože Spanring Biotehniška fakulteta Univerza E. Kardelja, Ljubljana Krekov trg 1, 61001 Ljubljana JUGOSLAVIJA

Dr. Ludwik Spiss Katedra Hodowli Roslin Lobzowska 24, A. Rolnicza Krakow POLAND

Dr. Veroslava Srejović kt Institut za biologiju, Prirodno-matematički fakultet. 34000 KRAGUJEVC JUGOSLAVIJA

Dr. Ivanka Stoeva Institute for Wheat and Sunflower 9520 Tolbuhin BULGARIA

Dr. Noriko Sugiyama Tokyo University of Agriculture, 1-1-1 Sakuraga-oka Setagaja-ku, Tokyo, 156, JAPAN

Dr. Saowanee Suputtitada Department of Genetics Faculty of Science Kasetsart University Bangkok 10900 THAILAND

Dr. I. M. Surikov Institut Puškin N. I. Vavilov V.I.R. Gercena, 44 190000 Leningrad SSSR

kr

is

Dr. Jan Szklarz Akademia Rolnicza Akademicka 15 Lublin POLSKA

Dr. Taras E. Ševčuk 1-6r Obščežitie sel'skohozjajstvennaja instituta ul. Timirjazeva 114, »A« gorod Kamenec-Podol'skij Hmelnickoi oblasti 281900 SSSR

Dr. Inayatullah Tahir 1–4nr Department of Botany University of Kashmir Srinagar – 190 006 Kashmir INDIA

Dr. Ljubov K. Taranenko Chabany, Kievskaja obl. Institut zemledelija 255205, USSR SSSR

Dr. Carlos shiguemi Tawara R. Dep. Nilson Ribas, 1045-CEP 86100 Jd. San Remo, Londrina, Paraná BRASIL

t

i

r

Dr. Barbara Thiem

Katedra Botaniki Farmaceutycznej Akademia Medyczna im. K. Marcinkowskiego, Wieniawskiego 1 61-712 Poznań POLAND

Dr. J. M. Thomas Ecole Nationale d-Ingenieurs des Travaux Agricoles F-21800 Quetigny FRANCE

Mr. Masahisa Tsuchiya

Managing Director Nikkoku Flour Milling Co., Ltd. 900 Minamichitosecho, Nagano 380 JAPAN

Dr. Eiji Tsuzuki Department of Agronomy Faculti of Agriculture Miyazaki University Miyazaki 880 JAPAN

University Microfilms International 300 North Zeeb Road, Box 91 Ann Arbor, Mi 48106 USA

Dr. Akio Ujihara air

ainoprs

2

Department of Crop Science and Plant Breeding Faculty of Agriculture Shinshu University Ina, Nagano-Ken JAPAN

James Udesky 1231 Lindenwood Drive Winnetka, Illinois 60093 USA

Vashnil

Centralnaja naučnaja sel'skohozjajstvennaja biblioteka Otdel meždunarodnogo knjigoobmena Orlikov per., 3. Moskva B-139 SSSR

ct Viniti

ar

i

A 219, Baltiskaja ul. 14 125219, Moskva SSSR

Blanka Vombergar-Dolinšek Bazoviška 10 62000 Maribor JUGOSLAVIJA

Bruce-D. Walker

2131 Red Deer Road Saskatoon, SASK CANADA

Dr. Maria Warcholowa Instytut Uprawy, Nawozenia i Gl. Csada Palacowa, 24-100 Pulawy POLAND

Dr. Lidija Weiss PUDOC General Foukesweg, P.B. 4 6700 AA WAGENINGEN THE NETHERLANDS

Dr. Giovanni Wittmer

Instituto Sperimentale per la Cerealicoltura Sezione operativa di Foggia S.S. n. 16-Km.675-CAS.Post. 1. 71100 FOGGIA ITALIA

Dr. Zdzislaw Zebrowski

Instytut Uprawy, Nawozenia i Gl. Osada Palacowa, 24-100 Pulawy POLAND

Teo Zor

SK Semenarna Gosposvetska c. 61000 LJUBLJANA JUGOSLAVIJA

Pawel Zorski

ul. Owcy-Orwicza 30-212 Kraków POLAND ar

i

S

i

Dr. Wang Zhongqing io Agriculture Academy of Inner Mongolia, HOHHOTO-CITY P.R. CHINA

Dr. T. Yabuya

Institut of Plant Breeding Faculty of Agriculture Miyazaki University Funatsuka 3-210, Miyazaki, Kyushu 880 JAPAN

Dr. Norio Yoshida

Ohsumi Branch, Kagoshima Agricultural Experim. Stn. 4938 Hosoyamada, Kushira-che Kimotsuki-gun, Kagoshima-ken JAPAN

Dr. Tomohiko Yoshida

Kyushu Agricultural **Eksperimental Station** Nishigoushi, Kumamoto, 861-11 JAPAN

Dr. Yoshiharu Yokoyama

NIPPON Flour Mills Co. Central Laboratory 2114-2 Nurumizu, Atsugi, 243 JAPAN

a

io Agriculture Academy of Inner Mongolia, HOHHOTO-CITY P.R. CHINA

Professional interest:

Genera and Species

- 1. Fagopyrum esculentum
- 2. Fagopyrum tataricum
- 3. Fagopyrum cymosum
- 4. Fagopyrum other species
- 5. Other genera
- 6. Interspecific and intergeneric

Usage

- a. Food
- b. Fodder
- c. Medicinal

Pests, diseases and weeds

- d. Animal
- e. Fungal
- f. Bacterial
- g. Virus
- h. Weed

Disciplines

Liu Yurui

- i. Agronomy
- j. Pathology
- k. Physiology
- I. Taxonomy m. Evolution
- **n.** Genetics and cytology
- o. Genetic resources
- p. Breeding
- q. Mating system
- r. Nutrition and chemical composition
- s. Seed production t. In vitro cultures